



The Journal of Anatomical Sciences

Email: journalofanatomicalsciences@gmail.com

J. Anat Sci 17(3) Sept.

Submitted: January 30th, 2026

Revised: July 2nd, 2026

Accepted: August 26th, 2026

DOI: <https://dx.doi.org/10.4314/jans.v17i3.2>

Preliminary Study of Nephroprotective Potentials of Ethanolic Leaf Extract of *Euphorbia Kamerunica* Against Aluminum Chloride-Induced Nephrotoxicity in Wistar Rats

Sanmi T. Ogunsanya^{1a}, Busayo J. Agboola², Abiodun K. Samuel³, Kehinde O. Adeniji¹, Stephen T. Adelodun¹, Oluseyi S. Fabiyi¹, Kelechi R. Onyema^{1b*}, Adetomiwa E. Adeogun⁴, Oluwafemi S. Olaniyi⁵

¹Department of Anatomy, Benjamin Carson Snr College of Health and Medical Sciences, Babcock University, Ilisan-Remo, Ogun State; ²Department of Biochemistry and Molecular Biology, Obafemi Awolowo University, Ile-Ife, Osun State; ³Biomedical Technology Research Group for Vulnerable Populations and the School of Health Science, Mae Fah Luang University, Muang, Chiang Rai 57100, Thailand; ⁴Department of Physiology, Benjamin Carson Snr College of Health and Medical Sciences, Babcock University, Ilisan-Remo, Ogun State; ⁵Department of Anatomy, University of Ilesa, Ilesa, Osun State, Nigeria

*Corresponding Author: Email: onyemak@babcock.edu.ng Tel: 08146679864
ORCID: 0000-0002-8088-4031^a; 0009-0003-1946-0226^b

ABSTRACT

Aluminium chloride (AlCl₃) is a widely used industrial reagent with well-documented nephrotoxic properties, capable of inducing significant oxidative stress, inflammation, and structural renal damage. This study evaluated the nephroprotective potential of the ethanolic leaf extract of *Euphorbia kamerunica* (EEEUK) in a Wistar rat model of AlCl₃-induced kidney injury, with emphasis on histoarchitectural and biochemical outcomes. Forty-two adult male Wistar rats (120–180 g) were randomized into six groups (n = 7): Control, AlCl₃ (200 mg/kg), EEEUK only (100 mg/kg), AlCl₃ + EEEUK (100 mg/kg), AlCl₃ + EEEUK (200 mg/kg), and AlCl₃ + donepezil (5 mg/kg) for 28 days (Oral administration of AlCl₃ for 14 days followed by oral administration of extract at varying doses for 14 days). At the end of the experiment, renal histomorphology, biochemical parameters, oxidative stress markers, and inflammatory indices were assessed. Phytochemical screening revealed abundant terpenoids and steroids, conferring significant antioxidant potency. Data obtained were analysed using statistical software (GraphPad Prism, 9.0). AlCl₃ administration induced severe renal pathology, including glomerular hypertrophy, tubular necrosis, vascular congestion, elevated serum creatinine, and heightened oxidative stress markers. EEEUK treatment dose-dependently reduced myeloperoxidase activity, superoxide anion formation, and peroxynitrite/hydroxyl radical generation, while restoring renal architecture and normalizing biochemical indices, with the 200 mg/kg dose demonstrating the most significant protective effect. EEEUK conferred significant nephroprotection against AlCl₃-induced renal toxicity through antioxidant, anti-inflammatory, and histomorphological preservation mechanisms, supporting its potential application in mitigating heavy-metal-related renal injury and warranting further investigation for therapeutic development.

Keywords: Aluminum chloride, *Euphorbia kamerunica*, nephrotoxicity, nephroprotection, oxidative stress

INTRODUCTION

Nephrotoxicity embodies a substantial clinical concern, delineated by the propensity of specific substances to inflict renal tissue injury and compromise normative kidney function¹. Organs that are especially susceptible to toxic insult because of their increased metabolic rate and concentrating properties include the kidneys, which are indispensable organs responsible for filtration, excretion, maintenance of electrolyte balance, and regulation of blood pressure².

Aluminum (Al), which is the third most common constituent in the crust of the earth, has become a major environmental and occupational health problem as it can be explained by the fact that it is extensively utilized in industry and that human beings are most

likely to come into contact with it³ as it is commonly used in a variety of industrial reactions including catalysis, water purification, and production⁴. Although useful, aluminium chloride poses significant health impacts, especially on the renal system, through mechanisms including oxidative stress, inflammatory pathways, and cellular damage⁵. Nephrotoxic effects of aluminium substances have been well documented both in experimental and clinical studies. The build-up of aluminium in the tissues of the kidney can trigger tubular cell death, necrosis of glomeruli, and worsening of renal function via the generation of reactive oxygen species (ROS) and disruptions in cellular homeostasis⁶. The vulnerability of the kidney to the effects of aluminium toxicity can be explained by the fact that the latter is involved not only in

aluminium excretion but also by the fact that the said metal tends to concentrate in the renal tissues ⁷.

Modern treatment options in nephrotoxicity induced by heavy metals mainly rely on chelation therapy combined with supportive treatment, which has a poor potential and often leads to detrimental side effects ⁸. This has led to a growing interest in the search for natural compounds that possess nephroprotective properties, especially those compounds that can be obtained from medicinal plants that have undergone proven safety and antioxidant activity ⁹.

Euphorbiaceae-type *Euphorbes*, such as *Euphorbia kamerunica* (commonly called cactus plant), are widespread succulent plants that occur throughout tropical and subtropical Africa ¹⁰. Traditional medicinal systems have recognized the potential of different *Euphorbia* species in the treatment of a range of disorders, including kidney diseases, inflammatory diseases, and oxidative stress-related diseases ¹¹. The plant contains a myriad of phytochemical products, which include diterpenoids, flavonoids, alkaloids, and phenolic compounds, all of which play a part in the therapeutic properties of the plant ¹². Recent phytochemical studies have revealed that *Euphorbia kamerunica* possesses high antioxidant, anti-inflammatory, and cytoprotective properties ¹³. Its bioactive compounds, more specifically terpenoids and steroids, have shown the ability to counteract free radicals and tame inflammatory signaling, as well as protect cellular bodies against oxidative injury ¹⁴. These properties highlight the possible therapeutic value of the plant in preventing and treating nephrotoxicity caused by heavy metals and other toxic agents.

Despite the documented biological activities of *Euphorbia kamerunica*, including antioxidant and anti-inflammatory properties, its nephroprotective potential against aluminium chloride-induced renal toxicity has not been investigated previously. The current study aimed to evaluate the nephroprotective properties of *Euphorbia kamerunica* ethanolic extract on the AlCl_3 -induced nephrotoxicity in Wistar rats with special focus on histomorphological changes, renal functional biochemical parameters, and variables of oxidative stress. Explaining the protective response mechanisms that exist in this medicinal plant can help add to the arsenal of knowledge on the design of innovative approaches to reverse renal injury caused by heavy metals.

MATERIALS AND METHODS

Plant material and extract preparation

Fresh leaves of *Euphorbia kamerunica* were obtained at the Okun-Owa village, Ijebu, Ogun State, Nigeria. Identification and authentication of the given specimen were done at the Ife Herbarium, Department of Botany, Obafemi Awolowo University, Ile-Ife, Nigeria, and a voucher specimen was deposited. The obtained leaves were washed and dried at ambient

temperature and finally crushed in a mechanical blender (Kenwood BL237, Kenwood, UK). The resulting powder (500g) was mixed with 80% and stirred with it over 24 hours at room temperature. The extract was then filtered on Whatman No.1 filter paper and concentrated with a rotary evaporator, which was operated at 40°C to produce the crude ethanolic extract of *Euphorbia kamerunica* (EEEUK) ¹⁵.

Experimental animals and ethics

The animal housing facility of Babcock University supplied forty-two male adult Wistar rats (body weight range of 120-180 g). Rats were housed in plastic cages under normal room conditions. Standard water and pellet feed were given *ad libitum*. This experiment was approved by the Babcock University Health Research and Ethics Committee (reference BUHREC 439/24) and conducted in accordance with the ethical standards of animal research ¹⁶. All procedures followed the Animal Research Reporting of In Vivo Experiments (ARRIVE) guidelines for reporting animal research ¹⁷.

Experimental design

After a period of one week of acclimatization in their plastic cages, the rats were randomly divided into six groups of seven animals each. Group A (Control) was treated with oral normal saline (2ml/kg) for 28 days. Group B (AlCl_3 alone) was orally administered 14 days of AlCl_3 (200mg/kg) for 14 days and normal saline. The ethanolic extract (EEEUK) at 100mg/kg was orally administered to Group C (EEEUK only) over 28 days. AlCl_3 (200 mg/kg) was added to Group D (AlCl_3 + EEEUK LD) for 14 days, followed by EEEUK (100 mg/kg) for 14 days. Group E (AlCl_3 +EEEUK HD) was treated with the addition of AlCl_3 (200 mg/kg) for 14 days, followed by EEEUK (200 mg/kg) for 14 days. Group F (AlCl_3 + Donepezil) was treated with 14 days of AlCl_3 (200 mg/kg) followed by 14 days of donepezil (5 mg/kg), thus used as the positive control. All therapies were administered orally through an oral cannula. The weights of the body were measured weekly during the time of the experiment ²⁰.

Sample collection and analysis

At the end (final day of administration) of the experimental period, animals were fasted overnight and sacrificed by cervical dislocation to minimize gastrointestinal content interference with tissue biochemistry, reduce anesthesia-related metabolic confounders, and ensure that cervical dislocation, a rapid, humane method causing immediate loss of consciousness through spinal cord severance, yields tissues with minimal stress-induced hormonal or metabolic artifacts, thereby preserving the histological and biochemical integrity of harvested organs. Blood samples were collected via cardiac puncture for biochemical analysis. Both kidneys were carefully dissected, excised, removed, weighed, and processed for histological and biochemical analyses.

Phytochemical analysis

Qualitative phytochemical screening was carried out according to the guidelines in an attempt to identify the existence of saponins, tannins, flavonoids, cardiac glycosides, terpenoids, steroids, phenols, alkaloids, and anthraquinones¹⁸. Antioxidant action of the ethanolic extract of the plant (EEEUK) was evaluated using 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay, using ascorbic acid as reference antioxidant¹⁹.

Evaluation of myeloperoxidase (MPO), reactive nitrogen species and superoxide dismutase (SOD) levels

Renal inflammation was assessed by measuring myeloperoxidase (MPO) activity, a well-established marker of neutrophil infiltration and oxidative-mediated tissue inflammation²⁰. Intracellular concentrations of reactive oxygen species (ROS) and reactive nitrogen species (RNS) were quantified via flow cytometry using the FACSCanto II system (Becton Dickinson, USA), following established fluorometric protocols^{21, 22}. Superoxide dismutase (SOD) activity was determined using the nitroblue tetrazolium (NBT) reduction method²³, which measures the enzyme's capacity to inhibit NBT reduction by superoxide radicals. Together, these complementary methodologies provided a comprehensive, multi-dimensional evaluation of the oxidative stress burden, antioxidant defense capacity, and inflammatory milieu within renal tissue, enabling robust interpretation of nephroprotective outcomes.

Histological analysis

Kidney tissues were fixed in 10% neutral buffered formalin and processed using the standard paraffin embedding technique. Sections of 5µm thickness were cut and stained with hematoxylin and eosin (H&E) for general histological examination, periodic acid-Schiff (PAS) for glycogen and basement membrane demonstration, and Masson's trichrome for collagen fiber visualization²⁴.

Statistical analysis

Data were expressed as mean ± standard error of mean (SEM). Statistical analysis was performed using GraphPad Prism 9.0. One-way analysis of variance (ANOVA) followed by Tukey's post-hoc test was used for multiple group comparisons. Statistical significance was set at $p < 0.05$ ²⁵

RESULTS

Phytochemical screening of *Euphorbia kamerunica*

Table 1 shows the phytochemical constituents of ethanolic extract of *Euphorbia kamerunica* (EEEUK); saponins, tannins, flavonoids, phenol, alkaloids, and anthraquinones are present, while terpenoids and

steroids are abundantly present. Cardiac glycosides are absent in EEEUK.

Organ weight changes

Kidney weight analysis revealed significant changes across groups. For the right kidney, there was a significant increase when control was compared with AlCl₃ only and EEEUK only groups. The AlCl₃ + EEEUK high dose groups showed a significant decrease compared to control. Left kidney weights showed similar patterns with significant differences across all groups. The EEEUK-only group had the highest kidney weight, while the AlCl₃ + EEEUK high dose group showed the lowest weight (Figure 1).

EEEUK mitigates renal inflammation damage by lowering myeloperoxidase function in AlCl₃-induced rats

Figure 3 illustrates a significant surge in MPO enzyme activities in the AlCl₃-induced group when compared with untreated animals. More importantly, the plant extract lowered the renal inflammatory response, as evidenced by a considerable decrease in MPO enzyme activity at 200 mg/kg when compared to the AlCl₃-induced group and the Donepezil-induced group (Figure 2).

Effect of EEEUK on superoxide dismutase activity

Superoxide dismutase (SOD) activity analysis revealed significant increases across all groups compared to control. The control group showed baseline SOD activity, while all experimental groups demonstrated elevated levels: AlCl₃ only, EEEUK only, AlCl₃ + EEEUK low dose, AlCl₃ + EEEUK high dose, and AlCl₃ + Donepezil. The progressive increase in SOD activity in treatment groups suggests enhanced antioxidant defense mechanisms (Figure 3).

Histological, histochemical and immunohistochemical examination of the kidney following AlCl₃ exposure

Histological examination of kidney sections across the four staining protocols revealed consistent patterns of AlCl₃-induced nephrotoxicity and dose-dependent EEEUK-mediated renoprotection. H&E staining demonstrated glomerular hypertrophy, vascular congestion, and tubular vacuolation in AlCl₃-treated rats, with progressive architectural recovery in EEEUK-treated groups (Figure 4). Masson Trichrome staining revealed marked interstitial fibrosis and collagen deposition following AlCl₃ administration, which was significantly attenuated by high-dose EEEUK treatment (Figure 5). PAS staining showed loss of brush borders and mesangial expansion in the AlCl₃ group, with marked restoration of PAS-positive reactivity in high-dose EEEUK-treated rats. Bcl-2 immunoreactivity was significantly diminished in

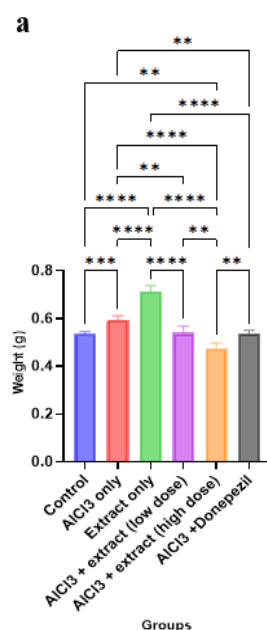
AlCl₃-treated kidneys, indicating increased apoptotic susceptibility (Figure 6), while EEEUK treatment dose-dependently upregulated Bcl-2 expression, confirming anti-apoptotic and renoprotective mechanisms (Figure 7). Donepezil-treated rats showed comparable protection across all staining protocols.

Table 1: Phytochemical screening of crude ethanolic extract

Test	Crude Ethanolic Extract
Saponins	+ve
Tanins	+ve
Flavonoids	+ve
Glycosides	-ve
Terpenoids	++ve
Steroids	++ve
Phenol	+ve
Alkaloids	+ve
Anthraquinone	+ve

s
NB; +ve: Present, ++ve: Abundantly present, -ve: Absent

RIGHT KIDNEY



LEFT KIDNEY

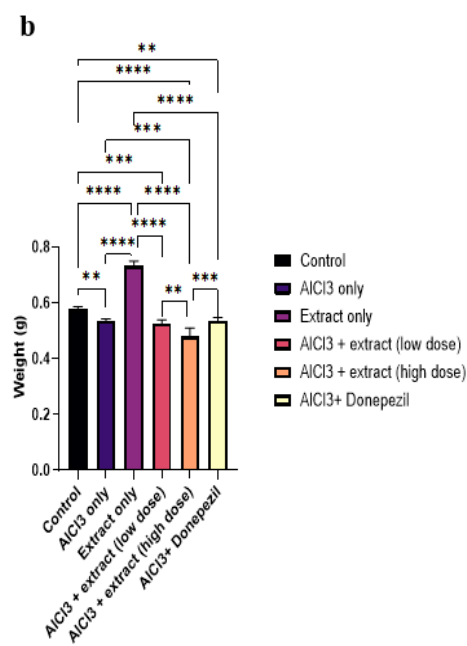


Figure 1 a and b: Bar chart showing the right and left kidneys' weight across all groups. Data are expressed as Mean $\pm$ SEM. **p<0.01, ***p<0.001, ****p<0.0001

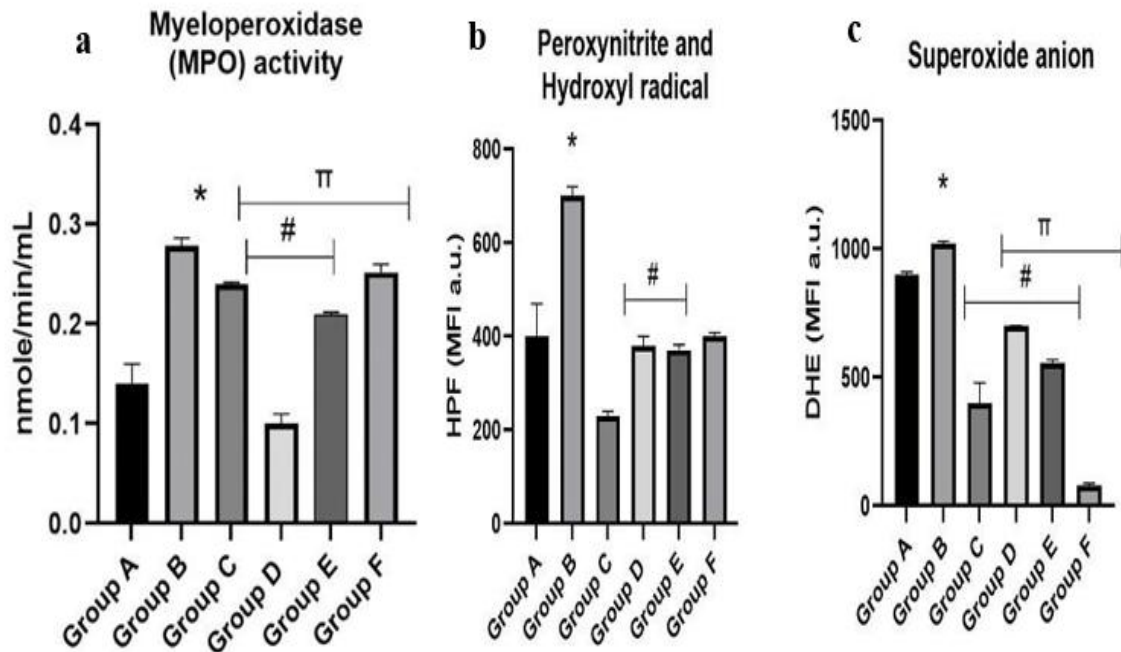


Figure 2 a-c: Bar charts showing EEUK mitigates renal inflammation by lowering (a) myeloperoxidase (MPO) (b) peroxynitrite and hydroxyl radical (c) superoxide anion production in AlCl_3 -induced Rats. Data are expressed as Mean $\pm$ SEM. * -denotes a substantial difference between the AlCl_3 group and the control group. # -denotes a substantial difference between the EEUK treatment cohorts and the AlCl_3 -induced group. While π -indicates a substantial difference between the positive control group and the EEUK treatment groups. Group A (Control): Administered normal saline orally; Group B (AlCl_3 only): Administered aluminium chloride (200 mg/kg body weight) orally; Group C (EEUK only): Administered EEUK (100 mg/kg body weight) orally; Group D (AlCl_3 + EEUK LD): Administered Aluminium chloride (200 mg/kg), followed by EEUK (100 mg/kg); Group E (AlCl_3 + EEUK HD): Administered aluminium chloride (200 mg/kg) for 14 days, followed by EEUK (200 mg/kg); Group F (AlCl_3 + Donepezil): Administered aluminium chloride (200 mg/kg) followed by donepezil (5 mg/kg) as a positive control.

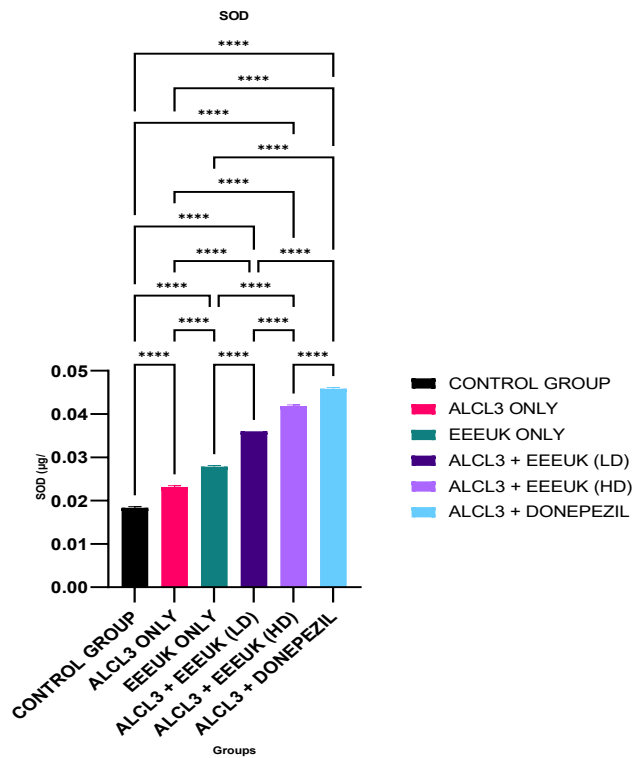


Figure 3: Bar charts showing superoxide dismutase (SOD) activity analysis revealed significant differences across all groups. Data are expressed as Mean $\pm$ SEM, ****- $p < 0.0001$

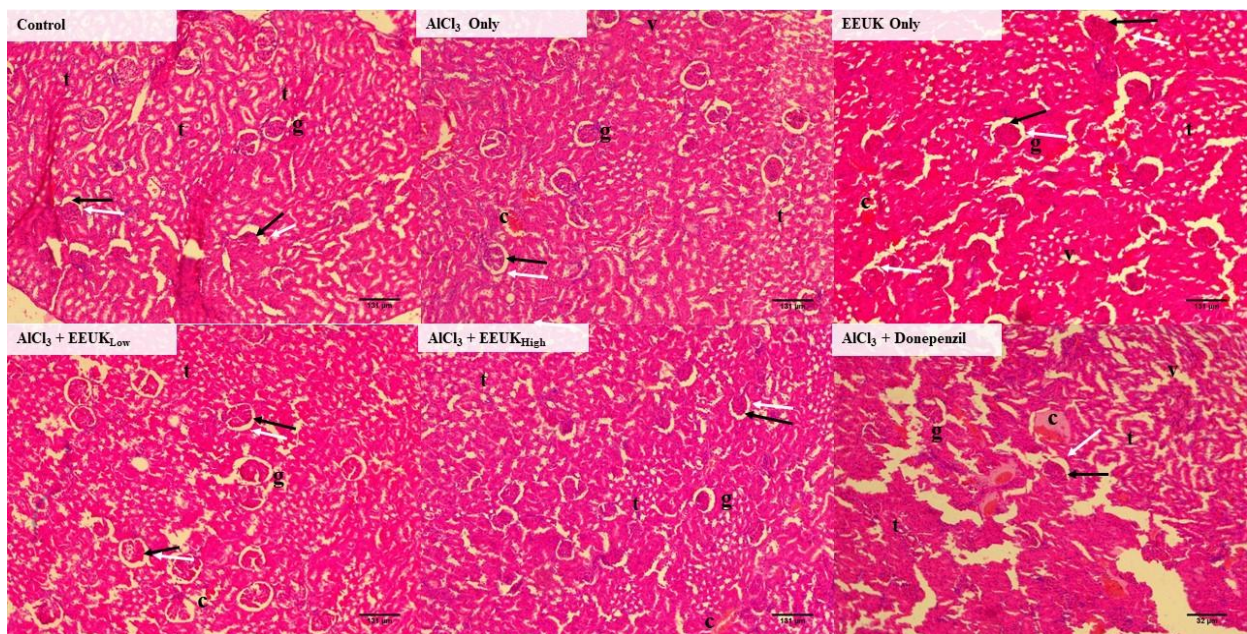


Figure 4: Photomicrograph of kidney section stained with H&E X400. The control group showed normal glomeruli (g), Bowman's capsule visceral (black arrow) and parietal (white arrow) layers, and proximal (p) and distal (d) convoluted tubules. AlCl_3 nephrotoxicity revealed enlarged glomeruli (g), congested blood vessels (c), and vacant areas between tubular cells and basal lamina (blue arrow). EEUK only showed nearly normal glomeruli (g) with slight change in convoluted tubules (d & p) and vacuolation (v). AlCl_3 + EEUK and AlCl_3 + Donepezil showed signs of recovery to normalcy in glomeruli (g), surrounding Bowman's capsule (black & white arrow), and tubules (d & p). Scale bar: 32 μm .

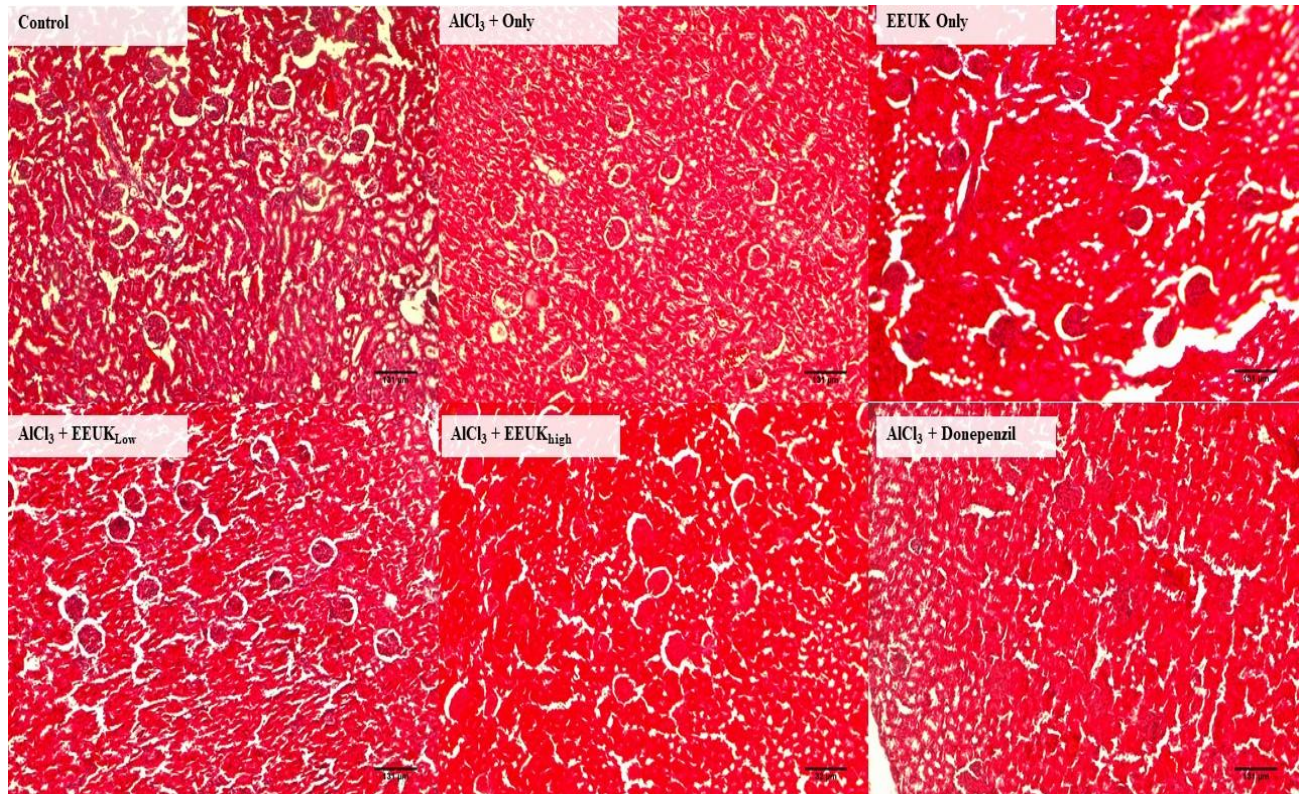


Figure 5: Photomicrographs of Masson Trichrome-stained kidney sections ($\times 400$); scale bar $32\ \mu\text{m}$) across groups: Control shows normal architecture; AlCl_3 -only exhibits severe tubular necrosis, collagen deposition (blue), and interstitial fibrosis; AlCl_3 +low-dose EEUK shows moderate improvement; AlCl_3 +high-dose EEUK demonstrates significant restoration of tubular integrity and reduced fibrosis; EEUK-only maintains normal histology; AlCl_3 +donepezil shows comparable renoprotection to high-dose EEUK treatment.

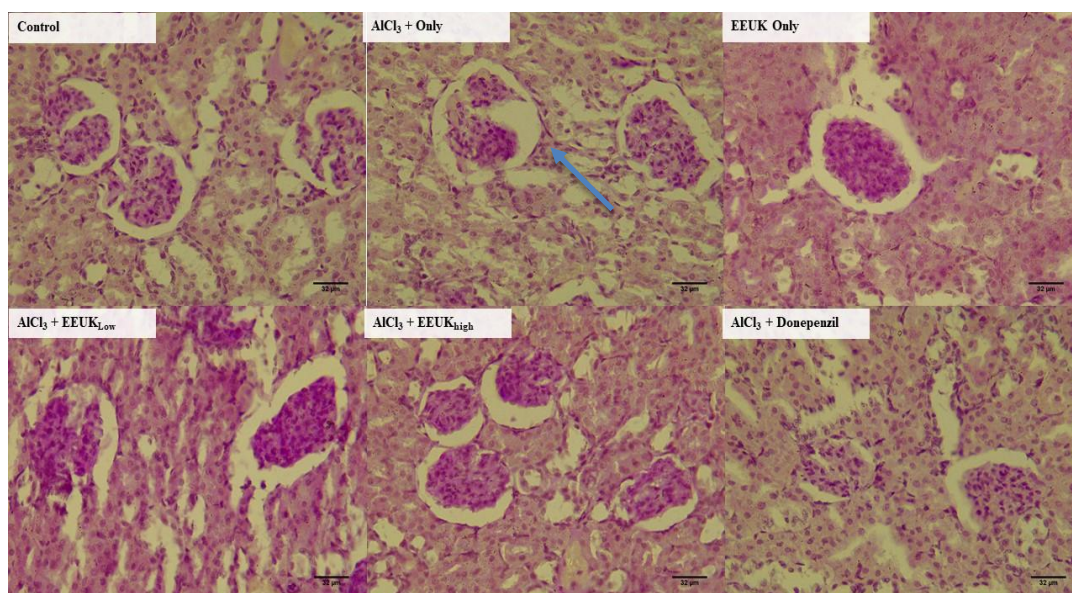


Figure 6: Photomicrographs of PAS-stained kidney sections ($\times 400$; scale bar $32\ \mu\text{m}$). Control has intact glomeruli and PAS-positive brush borders. AlCl_3 -only: loss of brush borders, tubular degeneration, mesangial PAS expansion (blue arrow). AlCl_3 +low-dose EEUK: partial restoration. AlCl_3 +high-dose EEUK: marked recovery with preserved PAS reactivity. EEUK-only: normal histology. AlCl_3 +donepezil: substantial improvement, comparable to high-dose EEUK, indicating protective effects.

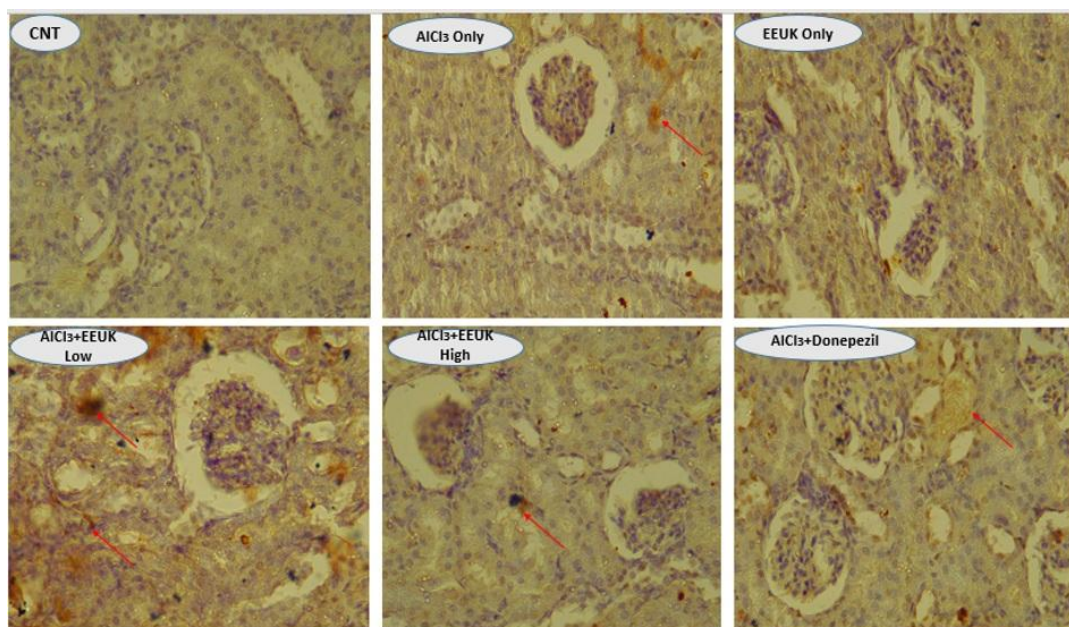


Figure 7: Photomicrographs showing BCL-2 immunoreactivity (brown staining) in kidney sections of Wistar rats (magnification $\times 400$). Control shows normal saline-treated rats displaying strong BCL-2 expression in renal tubular epithelial cells, indicating normal cellular homeostasis and anti-apoptotic activity. AlCl_3 only: Aluminum chloride (200 mg/kg) for 14 days followed by normal saline, showing significantly diminished BCL-2 immunoreactivity, suggesting aluminium-induced nephrotoxicity and increased apoptotic susceptibility. EEUK alone: EEUK (100mg/kg) for 28 days, demonstrating robust BCL-2 expression similar to controls. AlCl_3 + EEUK LD: Aluminum chloride followed by low-dose EEUK (100 mg/kg), showing moderate restoration of BCL-2 expression. AlCl_3 + EEUK HD: High-dose EEUK (200 mg/kg) treatment exhibiting marked BCL-2 upregulation and renoprotective effects. AlCl_3 + Donepezil: Positive control with donepezil (5 mg/kg) showing enhanced BCL-2 expression. Scale bar = 50 μm . Arrows indicate BCL-2-positive tubular cells.

DISCUSSION

The current study establishes strong nephroprotective properties of an ethanolic extract of *Euphoria kamerunica* over AlCl_3 -induced nephrotoxicity in Wistar rats. Such results offer good support to the therapeutic use of the medicinal plant in alleviating the effects of heavy metals on the kidneys by various protective mechanisms. EEUK possesses a variety of bioactive compounds, as phytochemical analysis showed that the compound has a significant proportion of terpenoids and steroids, which also play a role in its therapeutic effects. The high antioxidant property over ascorbic acid implies effective free-radical scavenging of the antioxidant, which is essential in the prevention of oxidative-stress-related cellular loss²⁶. These findings are in line with other literature that has shown the antioxidant effect of the species of the genus *Euphorbia* and its possible uses in therapy²⁷.

The administration of AlCl_3 was able to induce nephrotoxicity as observed in histopathological alterations and the modulation of markers of oxidative

stress. The renal injury (enlarged glomeruli, congested blood vessels, and tubular degeneration) is consistent with known pathways of aluminum-induced nephrotoxicity characterized by oxidative stress, inflammation, and dysfunction of cells²⁸. The effects of EEUK on oxidative stress and inflammation were observed via the cellular concentrations of MPO and production of reactive oxygen species (ROS) and reactive nitrogen species (RNS)²⁹. Both inflammation and oxidative stress are reported as signs of renal diseases^{30, 31}. The kidney, with its high metabolic needs and high level of mitochondrial density, is highly vulnerable to excessive ROS overproduction in stress situations³¹. According to Ebert *et al.*³⁰, acute inflammatory responses caused by the pathological stimuli increase the impact of oxidative stress and support the generation of ROS. The current research determined that the level of kidney myeloperoxidase activity and ROS were significantly elevated by AlCl_3 . Similar to the findings provided in the past, tissue injury and inflammation have been directly linked to MPO activity increase that is mostly due to inflammation^{33,34}. MPO enhances oxidative injury by promoting the conversion of hydrogen peroxide to

hypochlorite, thereby increasing cellular injury and the progression of inflammation³²⁻³⁵.

In addition to that, high levels of MPO have been shown to induce inducible nitric-oxide synthase (iNOS) and increase the production of nitric oxide (NO) [35]. NO, a crucial regulator of renal hemodynamics, is able to respond to ROS to form toxic peroxynitrite^{35, 36}. Under conditions of reduced intracellular concentrations, NO acts as a signaling molecule, controlling a number of physiological systems, including the endocrine system³⁷. However, high levels are widely known as one of the possible mediators of pro-inflammatory responses^{31,32,34}. This study proposes a potential increase in the concentration of NO in the kidneys of AlCl₃-treated rats, causing nitrosative stress because of the formation of peroxynitrite, having a potential to cause tissue damage^{34,35}. However, previous studies suggest that the species of *Euphorbia* alleviate apoptotic activity caused by H₂O₂³⁸. The present research showed that EEEUK reduced the initial increase in inflammatory biomarkers produced by AlCl₃, and the effects were especially significant in lowering the level of MPO, ROS, and RNS in renal cells of experimental units. The results are consistent with the earlier studies that have indicated the antithrombotic and anti-inflammatory effects of methanolic leaf extracts of the plant species of the genus *Euphorbia* on the kidney³⁹. The positive impact of the EEEUK could be attributed to its antioxidant capability that fights oxidative damage⁴⁰. EEEUK can also include bioactive molecules, including alkanols and terpenoids, which react with the active site of MPO and counteract its actions⁴¹.

The increase in SOD activity in AlCl₃-treated animals is a compensatory mechanism for oxidative stress. SOD is an essential antioxidant enzyme that counteracts superoxide radicals, and its up-regulation points to the efforts of the organism to overcome the effects of aluminum-related oxidative damage⁴². The incremental upsurge of SOD activity in EEEUK-treated groups indicates the improvement of the antioxidant defense systems, which favors the protective effects of the plant. EEEUK exhibited nephroprotective characteristics using several parameters. The dose-dependent improvement of renal architecture was observed by the histopathological examination, whereby the high dose group exhibited better restoration of glomerular architecture and less tubular damage. Basement-membrane integrity, which is evidenced by PAS staining, is a sign of protection against structural damage caused by aluminum. The decreased accumulation of collagen in the trichrome staining technique by Masson indicates the inhibition of the fibrotic alterations that come with chronic nephrotoxicity.

The pathways that have led to the described nephroprotective activity of EEEUK are probably multifarious. Antioxidant activity, anti-inflammatory effects, and cellular protection may be attributed to the high phytochemical content of the plant, with terpenoids and steroids being the main ones⁴³. Flavonoids and phenolic compounds found in the extract are characterized by the power to eliminate free radicals, alter effects of inflammation, and protect the cell membranes against oxidative injury⁴⁴. The dose-dependent effects observed in this study show that an increase in bioactive compound concentration offers greater protection. Histopathological preservation and antioxidant enzyme activity were also more favorable in the high-dose (200mg/kg) group than the low one, which shows that there is a therapeutic limit of nephroprotection. The therapeutic potential of this plant extract is justified by the similarity of its effectiveness to the clinically administered neuroprotective agent, donepezil. However, the natural origin of EEEUK could have benefits in safety profile and fewer adverse effects than synthetic drugs⁴⁵.

The aluminum chloride considerably decreased the expression of BCL-2 in the renal tubular cells, which implies an increase in the susceptibility to apoptosis and nephrotoxicity, as reported earlier. The anti-apoptotic protein BCL-2 is the one that preserves mitochondrial membrane integrity and inhibits cytochrome c leakage^{46,47}. There was a dose-dependent restoration of BCL-2 immunoreactivity by EEEUK treatment, which implicated Renoprotective pathways via apoptosis suppression. The high-dose EEEUK sample exhibited similar efficacy to donepezil, suggesting the presence of powerful anti-apoptotic activities that could be due to the presence of antioxidant phytochemicals⁴⁸. The research results are consistent with those that indicate that natural compounds reduce organ damage caused by aluminum by increasing survival proteins⁴⁹. BCL 2 replacement indicates a positive association with renal architecture, which is in favor of the therapeutic effect of EEEUK against metal-induced nephrotoxicity.

Limitations of the study are rather limited period of treatment and acute nephroprotective effects. Research in the future should focus on long-term preventive effects, dose route optimization and bioactive compounds that are causing a protective effect on the kidney. Also, mechanistic research on the molecular pathways of EEEUK protective actions would be informative to the development of a therapeutic intervention.

CONCLUSIONS

This study has established that an ethanolic extract of *Euphorbia kamerunica* has considerable nephroprotective properties in nephrotoxicity induced by AlCl₃ in Wistar rats. The therapeutic potential of *Euphorbia kamerunica* in the prevention and

treatment of nephrotoxicity in human beings will require a clinical trial to validate its therapeutic potential.

REFERENCES

- Perazella MA. Renal vulnerability to drug toxicity. *Clin J Am Soc Nephrol*. 2009;4(7):1275-83.
- Alwiyah F, Rudiyanto W, Anggraini DI, Windarti I. Anatomy and physiology of the kidney: literature review. *Med Prof J Lampung*. 2024;14(2):285-9.
- Krewski D, Yokel RA, Nieboer E, Borchelt D, Cohen J, Harry J, *et al*. Human health risk assessment for aluminum, aluminum oxide, and aluminum hydroxide. *J Toxicol Environ Health B Crit Rev*. 2007;10 Suppl 1:1-269.
- Calkins S. Tea waste as a green method in removing aluminum from drinking water [dissertation]. Norton (MA): Wheaton College; 2022.
- Wei H, Li D, Luo Y, Wang Y, Lin E, Wei X. Aluminum exposure induces nephrotoxicity via fibrosis and apoptosis through the TGF- β 1/Smads pathway in vivo and in vitro. *Ecotoxicol Environ Saf*. 2023;249:114422.
- Alfrey AC, LeGendre GR, Kaehny WD. The dialysis encephalopathy syndrome: possible aluminum intoxication. *N Engl J Med*. 1976;294(4):184-8.
- Frost RW, Lasseter KC, Noe AJ, Shamblen EC, Lettieri JT. Effects of aluminum hydroxide and calcium carbonate antacids on the bioavailability of ciprofloxacin. *Antimicrob Agents Chemother*. 1992;36(4):830-2.
- Naughton CA. Drug-induced nephrotoxicity. *Am Fam Physician*. 2008;78(6):743-50.
- Welz AN, Emberger-Klein A, Menrad K. Why people use herbal medicine: insights from a focus-group study in Germany. *BMC Complement Altern Med*. 2018;18(1):92.
- Burkill HM. *The Useful Plants of West Tropical Africa*. 2nd ed. Kew: Royal Botanic Gardens; 1994–2000
- Tsobou R, Mapongmetsem PM, Van Damme P. Medicinal Plants Used for Treating Reproductive Health Care Problems in Cameroon, Central Africa¹. *Econ Bot*. 2016;70:145-159
- Majid M, Khan MR, Shah NA, Ul Haq I, Farooq MA, Ullah S, *et al*. Studies on phytochemical, antioxidant, anti-inflammatory and analgesic activities of *Euphorbia dracunculoides*. *BMC Complement Altern Med*. 2015;15:349
- Pharmacological significance, medicinal use, and toxicity of extracted and isolated compounds from *Euphorbia* species found in Southern Africa: a review. *Plants (Basel)*. 2025;14(3):469.
- Zhao JX, Shi SS, Sheng L, Li J, Yue JM. Terpenoids and steroids from *Euphorbia hypericifolia*. *Nat Prod Commun*. 2015;10:2049-2052.
- Abubakar AR, Haque M. Preparation of medicinal plants: basic extraction and fractionation procedures for experimental purposes. *J Pharm Bioallied Sci*. 2020;12(1):1-10.
- National Research Council. Guide for the care and use of laboratory animals. 8th ed. Washington (DC): The National Academies Press; 2011.
- Kilkenny C, Browne WJ, Cuthill IC, Emerson M, Altman DG. Improving bioscience research reporting: the ARRIVE guidelines for reporting animal research. *PLoS Biol*. 2010;8(6):e1000412.
- Harborne JB. *Phytochemical methods: a guide to modern techniques of plant analysis*. 3rd ed. London: Chapman and Hall; 1998.
- Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. *LWT Food Sci Technol*. 1995;28(1):25-30.
- Bradley PP, Christensen RD, Rothstein G. Cellular and extracellular myeloperoxidase in pyogenic inflammation. *Blood*. 1982;60(3):618-22.
- Dias AT, Rodrigues BP, Porto ML, Gava AL, Balarini CM, Freitas FP, *et al*. Sildenafil ameliorates oxidative stress and DNA damage in the stenotic kidneys in mice with renovascular hypertension. *J Transl Med*. 2014;12(1):35.
- Tonini CL, Campagnaro BP, Louro LP, Pereira TM, Vasquez EC, Meyrelles SS. Effects of aging and hypercholesterolemia on oxidative stress and DNA damage in bone marrow mononuclear cells in apolipoprotein E-deficient mice. *Int J Mol Sci*. 2013;14(2):3325-42.
- Bancroft JD, Gamble M. *Theory and practice of histological techniques*. 6th ed. Edinburgh: Churchill Livingstone; 2008.
- Beauchamp C, Fridovich I. Superoxide dismutase: improved assays and an assay applicable to acrylamide gels. *Anal Biochem*. 1971;44(1):276-87.
- Field A. *Discovering statistics using SPSS*. 4th ed. London: SAGE Publications; 2013.
- Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: an overview. *ScientificWorldJournal*. 2013;2013:162750.

27. Tungmunthum D, Thongboonyou A, Pholboon A, Yangsabai A. Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: an overview. *Medicines (Basel)*. 2018;5(3):93.
28. Verstraeten SV, Aimo L, Oteiza PI. Aluminum and lead: molecular mechanisms of brain toxicity. *Arch Toxicol*. 2008;82(11):789-802.
29. Malle E, Buch T, Grone HJ. Myeloperoxidase in kidney disease. *Kidney Int*. 2003;64(6):1956-67.
30. Ebert T, Neytchev O, Witas A, Kublickiene K, Stenvinkel P, Shiels PG. Inflammation and oxidative stress in chronic kidney disease and dialysis patients. *Antioxid Redox Signal*. 2021;35(17):1426-48.
31. Fernandes SM, Watanabe M, Vattimo MD. Inflammation: improving understanding to prevent or ameliorate kidney diseases. *J Venom Anim Toxins Incl Trop Dis*. 2021;27:e20200162.
32. Aranda-Rivera AK, Cruz-Gregorio A, Aparicio-Trejo OE, Pedraza-Chaverri J. Mitochondrial redox signaling and oxidative stress in kidney diseases. *Biomolecules*. 2021;11(8):1144.
33. Ismail HM, Ahmed SA, Alsaedi AM, Almaramhy WH, Alraddadi MK, Albadrani MS, *et al*. Reactive oxygen and nitrogen species in myocardial infarction: mechanistic insights and clinical correlations. *Med Sci (Basel)*. 2025;13(3):152.
34. Zhang X, Zhu Y, Xiong Z, Xie W, Shao M, Liu Z. Broad-spectrum ROS/RNS scavenging catalase-loaded microreactors for effective oral treatment of inflammatory bowel diseases. *Small*. 2025;21(23):2501341.
35. Bahadoran Z, Carlström M, Mirmiran P, Ghasemi A. Nitric oxide: to be or not to be an endocrine hormone? *Acta Physiol (Oxf)*. 2020;229(1):e13443.
36. Athmouni K, Belhaj D, El Feki A, Ayadi H. Optimization, antioxidant potential, modulatory effect and anti-apoptotic action of *Euphorbia bivonae* polysaccharides on hydrogen peroxide-induced toxicity in human embryonic kidney cells HEK293. *Int J Biol Macromol*. 2018;116:482-91.
37. Rahman MS, Rana S, Islam AA. Antithrombotic and anti-inflammatory activities of leaf methanolic extract of *Euphorbia hirta* Lin. *Int J Complement Alt Med*. 2019;12(4):154-62.
38. Barla A, Öztürk M, Kültür Ş, Öksüz S. Screening of antioxidant activity of three *Euphorbia* species from Turkey. *Fitoterapia*. 2007;78(6):423-5.
39. Ogunnusi TA, Oso BA, Dosumu OO. Isolation and antibacterial activity of triterpenes from *Euphorbia kamerunica* Pax. *Int J Biol Chem Sci*. 2010;4(1):158-67.
40. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. *World Allergy Organ J*. 2012;5(1):9-19.
41. Adebayo SA, Dzoyem JP, Shai LJ, Eloff JN. The anti-inflammatory and antioxidant activity of 25 plant species used traditionally to treat pain in southern Africa. *BMC Complement Altern Med*. 2015;15:159.
42. Spencer JP. The impact of fruit flavonoids on memory and cognition. *Br J Nutr*. 2010;104 Suppl 3:S40-7.
43. Mahli A, Koch A, Czech B, Peterburs P, Lechel A, Haunschild J, *et al*. Hepatoprotective effect of oral application of a silymarin extract in carbon tetrachloride-induced hepatotoxicity in rats. *Br J Pharmacol*. 2015;172(1):7-18.
44. Yousef MI, Salama AF, Shalaby MA, El-Gendy KS. Aluminum-induced deterioration in renal function and structure. *Toxicol Ind Health*. 2019;35(7):464-75.
45. Sharma V, Sharma A, Kansal L. Neuroprotective potential of herbal extracts against aluminum-induced toxicity. *J Ethnopharmacol*. 2020;252:112598.
46. Czabotar PE, Lessene G, Strasser A, Adams JM. Control of apoptosis by the BCL-2 protein family: implications for physiology and therapy. *Nat Rev Mol Cell Biol*. 2014;15(1):49-63.
47. Abdel-Daim MM, Abushouk AI, Donia T, Alarifi S, Alkahtani S, Aleya L, *et al*. Protective effects of thymoquinone against aluminum-induced nephrotoxicity. *Toxicol Mech Methods*. 2018;28(6):451-8.